

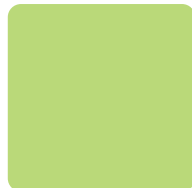


Warsaw University of Technology

Faculty of Power
and Aeronautical Engineering



XXVI FLUID MECHANICS CONFERENCE



10-13 September 2024 - Warsaw, Poland

BOOK OF ABSTRACTS

Editors: S. Gepner, S. Kubacki, T. Waclawczyk

J. A. M. Méndez, B. Dorneanu, H. Schmidt, H. Arellano-García: Revisiting homogeneous modeling with volume averaging theory: structured catalysts for steam reforming and CO₂ methanation	52
K. Wawrzak, A. Wawrzak, A. Bogusławski, A. Tyliczszak: A high-order LES of a flow in complex geometry	54
J. Piechna, M. Szudarek, A. Piechna: Numerical modelling of a rotary shock wave compression heat engine with a rotating detonation wave combustion chamber	56
S. Telyma, O. Oliynyk: Some theoretical problems of creation the mathematic model of joint treatment of wastewaters with organic contaminants and nitrogen compounds by method of biofiltration	58
COMPUTATIONAL FLUID DYNAMICS	60
A. Kajzer: Isotropy of numerical errors in the context of implicit large eddy simulation	61
J. Fabisiak, S. Gepner: Quantification of laminar mixing efficiency using ‘strange eigenmodes’ approach	63
P. Radomski, D. Kreft, P. Ziółkowski, I. Mukha, J. Zieliński, D. Mikielwicz: Heat transfer of laser-illuminated gold nanorod platforms distributed in a flow germicidal chamber	65
M. Marek: Numerical modelling of gas flow in random packed bed with a helical flow deflector	67
S. Koval, N. Dimitrieva: Numerical simulation of two-phase flow in OpenFOAM software	69
S. Motoki, G. Kawahara: Dissimilar heat transfer enhancement in pipe flow with deep axial grooves	71
T. Bodnar: Numerical evaluation of mass diffusive compressible fluids flows models	73
A. Couvez, S. Gyuran, N. Leterrier, P. Omnes, E. Saikali: Code coupling for the Tube Support Plate clogging in steam generators	75
P. Niegodajew: Numerical investigation of Air Flow within a Human Nasal Cavity	77
V. Oliynik, T. Batutina: Semiempirical model of the acoustics of a supersonic jet upon collision with a perpendicular wall	79
J. Malet, R. Ploix, E. Géhin: Step-by-step CFD validation of turbulent particle transport and deposition in industrial bends	81
J. Gałecki, J. Szumbariski: High performance least-squares spectral/hp element method solvers for fluid dynamics problems	83
B. Kopiczak, K. Karbowski, K. Nering, Z. Malecha, R. Chrzan, J. Gawlik, A. Sucherska, J. Szaleniec, J. Karbowski: Application of CFD airflows to aid nasal obstruction diagnosis	85
I. Gorban: Application of the model of trapped vortices to the control of flow around a bridge pear	87
N. Yurchenko, P. Vynogradskyy, R. Pavlovskyy: Improvement of the aerodynamic performance using the developed method of energy-efficient flow control	89
EXPERIMENTAL METHODS	91
P. Korczyk., T. Kurniawan, S. Błoński, B. Kupikowska-Stobba: Integrated Approaches in Microfluidic Design for Enhanced Droplet Manipulation and Biological Insights	92
K. Bukowski, Ł. Klotz: Influence of micro- and mesoscale on the permeability characteristics of 3D printed porous objects	94
D. Duda, V. Yanovych, V. Uruba: PIV measurement of model nuclear fuel rod bundle	96
T. Kowalewski: Experimental Challenges of Nano and Microfluidics	98

Integrated Approaches in Microfluidic Design for Enhanced Droplet Manipulation and Biological Insights

P M Korczyk¹, T Kurniawan¹, S Błoński¹, B Kupikowska-Stobba¹

¹Institute of Fundamental Technological Research, Polish Academy of Sciences, ul. Pawińskiego 5B, 02-106 Warszawa, Poland

E-mail: piotr.korczyk@ippt.pan.pl

Abstract. The Institute of Fundamental Technological Research's Microfluidic Laboratory is focused on enhancing the accuracy and practical use of microfluidic methods for chemical and biological studies, as well as creating tailored microfluidic instruments to address specific biological research needs. In this document, we present a few of our latest projects.

Keywords: Micro-, Nano- and Bio-flows, Multi-phase Flows, Droplets

1. Introduction

Microfluidics has rapidly evolved from its inception, becoming a vital interdisciplinary field that spans fluid mechanics at microscales and has myriad uses in biology, chemistry, and diagnostics [1]. Its allure lies in the ability to precisely control and manipulate fluid flows, achieved through the tiny dimensions of microchannels, the smooth, laminar flow characteristics [2], and the significant influence of surface tension in flows involving different phases [3]. Our research focuses on both single and multiphase flows.

2. Droplet microfluidics

Using more than one immiscible phase allows for the controlled formation [4] and manipulation of droplets in microfluidic channels. Each droplet can be a miniature reactor containing samples, reagents, or biological components.

Our research delves into the basic principles of two-phase flows within microchannels [4,5]. Utilizing the insights gained, we craft innovative microfluidic designs, such as for the passive handling of droplets [6,7] and sequential logic devices for meticulous droplet management [7,8]. These advancements allow for the execution of complex procedures intricately encoded within the layout of our microfluidic systems.

Additionally, we employ digital algorithms to precisely adjust concentrations by selectively merging and equally dividing droplets, enhancing our processes' accuracy, repeatability, and adaptability [9].

3. Applications in biomedical research

Confined geometry of microfluidic chambers and superior flow control renders this technology suitable for mimicking physiological conditions for culturing cells [10]. One of our lab's primary objectives is to tailor microfluidic devices to meet specific biological research needs. A notable achievement is developing a microfluidic system that precisely controls the generation of tension gradients through the deformation of epithelial layers, enabling a detailed study of tissue mechanics, including strain and curvature effects on epithelial responses.

In collaboration with the University Grenoble Alpes in France, this system's application has provided insights into how curvature influences the propagation of calcium waves caused by folding on short timescales and affects gene expression spatially over more extended periods [11]. Our findings reveal that gradients in cell shape and the mechanical stresses they induce lead to distinct biochemical responses across the tissue layer, offering new perspectives on cell differentiation mechanisms during tissue development.

Another example is a device developed in collaboration with the University of Oxford, designed to study erythrocytes' oxygen release rate [12]. Our microfluidic system with the medium exchange chamber was applied for an experimental method to monitor the oxygen flow in individual red blood cells, combining ultrarapid solution switching to manipulate gas tension with single-cell O₂ saturation fluorescence microscopy.

Recently, this approach has been used to investigate human kidneys perfused with stored blood during transplantation; the respiratory rate of the organ was measured [13]. The study challenges the conventional definition of oxygen delivery based on blood flow and oxygen content, highlighting its inadequate representation of blood efficiency in tissue oxygenation. However, the research uncovered a robust correlation between monitored kidney respiration and erythrocytes' oxygen release rate.

References

- [1] G.M. Whitesides, The origins and the future of microfluidics, *Nature* 442 (2006) 368–373.
- [2] K.W. Oh, K. Lee, B. Ahn, E.P. Furlani, Design of pressure-driven microfluidic networks using electric circuit analogy, *Lab Chip* 12 (2012) 515–545. <https://doi.org/10.1039/C2LC20799K>.
- [3] C.N. Baroud, F. Gallaire, R. Danga, Dynamics of microfluidic droplets, *Lab Chip* 10 (2010) 2032–2045.
- [4] P.M. Korczyk, V. van Steijn, S. Blonski, D. Zaremba, D.A. Beattie, P. Garstecki, Accounting for corner flow unifies the understanding of droplet formation in microfluidic channels, *Nature Communications* 10 (2019) 2528.
- [5] T. Kurniawan, M. Sahebdivani, D. Zaremba, S. Blonski, P. Garstecki, V. van Steijn, P.M. Korczyk, Formation of droplets in microfluidic cross-junctions at small capillary numbers: Breakdown of the classical squeezing regime, *Chemical Engineering Journal* 474 (2023) 145601. <https://doi.org/10.1016/j.cej.2023.145601>.
- [6] P.M. Korczyk, L. Derzsi, S. Jakiela, P. Garstecki, Microfluidic traps for hard-wired operations on droplets, *Lab Chip* 13 (2013) 4096–4102. <https://doi.org/10.1039/C3LC50347J>.
- [7] D. Zaremba, S. Blonski, M. Jachimek, M.J. Marijnissen, S. Jakiela, P.M. Korczyk, Investigations of modular microfluidic geometries for passive manipulations on droplets, *Bull. Pol. Acad. Sci.-Tech. Sci.* 66 (2018) 139–149.
- [8] D. Zaremba, S. Błonski, P.M. Korczyk, Integration of capillary–hydrodynamic logic circuitries for built-in control over multiple droplets in microfluidic networks, *Lab Chip* 21 (2021) 1771–1778. <https://doi.org/10.1039/D0LC00900H>.
- [9] D. Zaremba, S. Blonski, P.M. Korczyk, Concentration on demand – A microfluidic system for precise adjustment of the content of single droplets, *Chemical Engineering Journal* 430 (2022) 132935.
- [10] D.N. Breslauer, P.J. Lee, L.P. Lee, Microfluidics-based systems biology, *Mol. BioSyst.* 2 (2006) 97–112.
- [11] S. Blonski, J. Aureille, S. Badawi, D. Zaremba, L. Pernet, A. Grichine, S. Fraboulet, P.M. Korczyk, P. Recho, C. Guilluy, M.E. Dolega, Direction of epithelial folding defines impact of mechanical forces on epithelial state, *Developmental Cell* 56 (2021) 3222–3234.e6. <https://doi.org/10.1016/j.devcel.2021.11.008>.
- [12] J. Rabcuka, S. Blonski, A. Meli, S. Sowemimo-Coker, D. Zaremba, D. Stephenson, M. Dzieciatkowska, D. Nerguizian, R. Cardigan, P.M. Korczyk, P.A. Smethurst, A. D'Alessandro, P. Swietach, Metabolic reprogramming under hypoxic storage preserves faster oxygen unloading from stored red blood cells, *Blood Advances* 6 (2022) 5415–5428. <https://doi.org/10.1182/bloodadvances.2022007774>.
- [13] R. Dumbill, J. Rabcuka, J. Fallon, S. Knight, J. Hunter, D. Joyce, J. Barrett, M. Ellen, A. Weissenbacher, T. Kurniawan, S. Blonski, P.M. Korczyk, R. Ploeg, C. Coussios, P. Friend, P. Swietach, Impaired O₂ unloading from stored blood results in diffusion-limited O₂ release at tissues: evidence from human kidneys, *Blood* 143 (2024) 721–733. <https://doi.org/10.1182/blood.2023022385>.